

#### Multistep protocol of immunoassays requires automated diagnostics device in remote areas

#### RESULTS

- Fully enclosed microfluidic paper-based analytical devices fabricated by fluorocarbon (pentafluoroethane (PFE)) plasma deposition followed by  $O_2$  plasma etching.
- Flow channels with different wettability can be formed by varying  $O_2$  plasma etch time
- Flow channels protected by hydrophobized paper layers to **inhibit contamination**
- Channels sealed with adhesive tape to prevent sample evaporation
- Device operation demonstrated by three reagents with different pH values sequentially **reacting** with indicator at detection zone

# CONCLUSION

- Proposed µ-PADs make possible automated multistep protocol assays (e.g. ELISA) to be employed for low-cost disease diagnostics in remote locations
- Fully enclose channels can be easily packaged using adhesive tape to eliminate need for device packaging, thus decreasing overall device cost

# REFERENCES

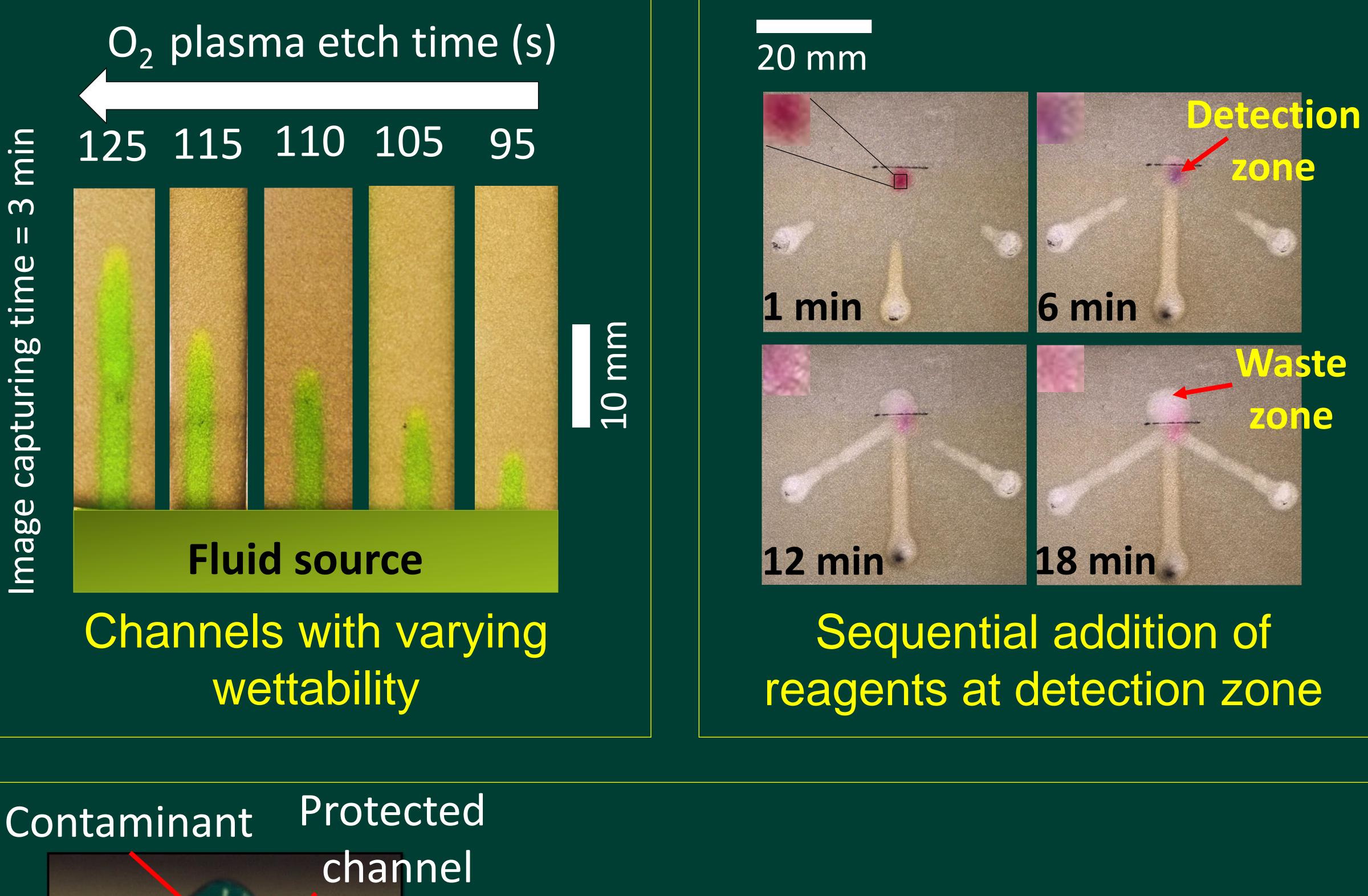
- 1. https://www.cusabio.com/c-15109.html
- 2. Raj et al., Sensors and Actuators B: Chemical, 2020
- 3. Raj et al., Lab on a Chip, 2019

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# **INCORPORATING FLOW CONTROL FUNCTIONALITY** IN MICROFLUIDIC PAPER-BASED ANALYTICAL DEVICES USING PLASMA PROCESS

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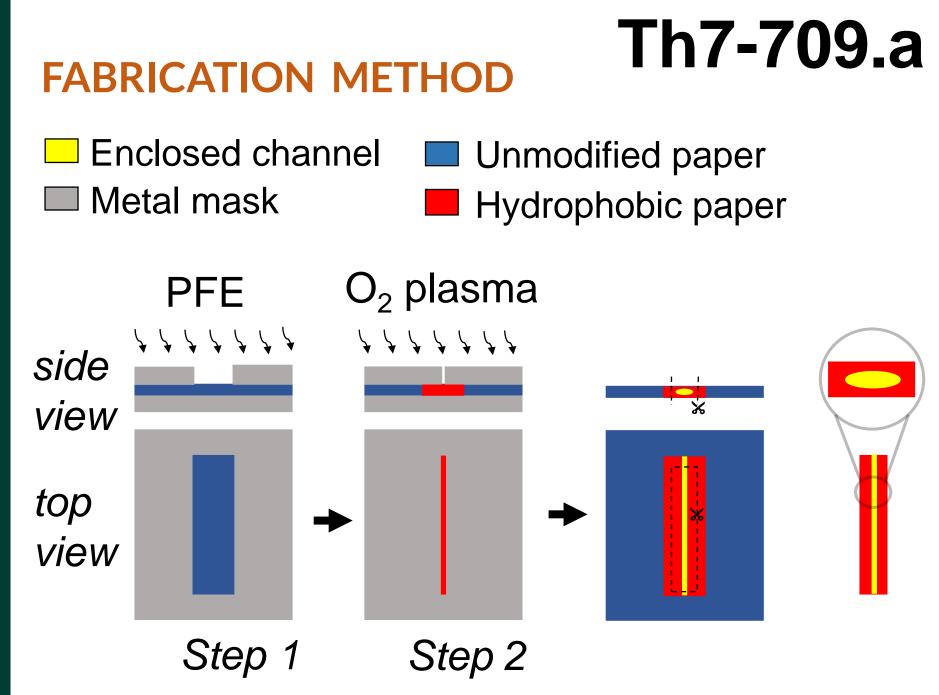


Cross-sectional image

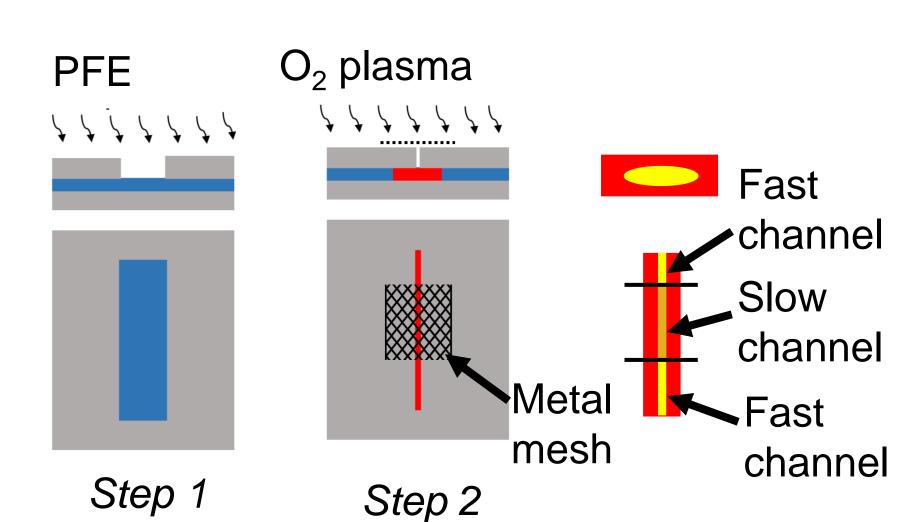
1 mm

Take home: The proposed paper-based device design can automate multistep protocol assays thereby making diagnostic tests such as ELISA a viable option for low-cost disease diagnostics in remote areas

Fully enclosed channels for inhibition of contamination and evaporation

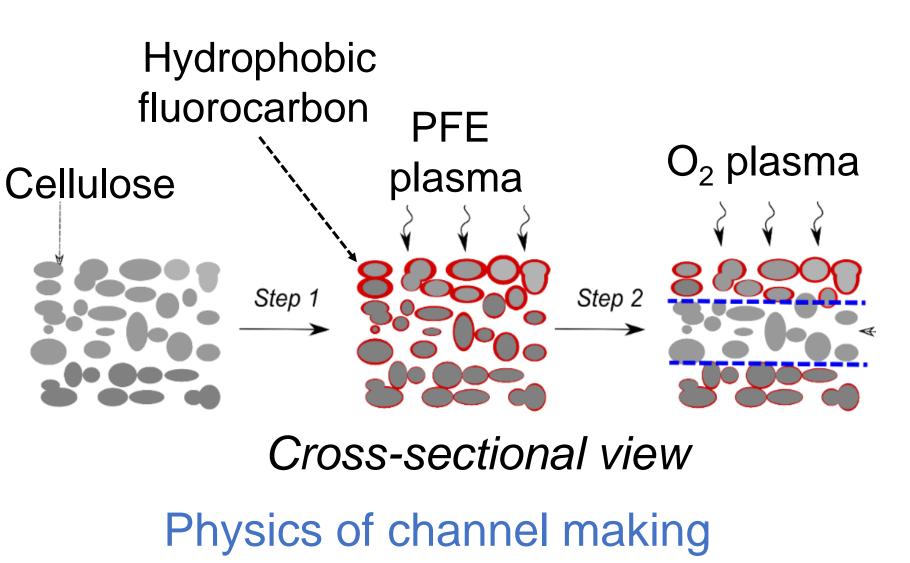


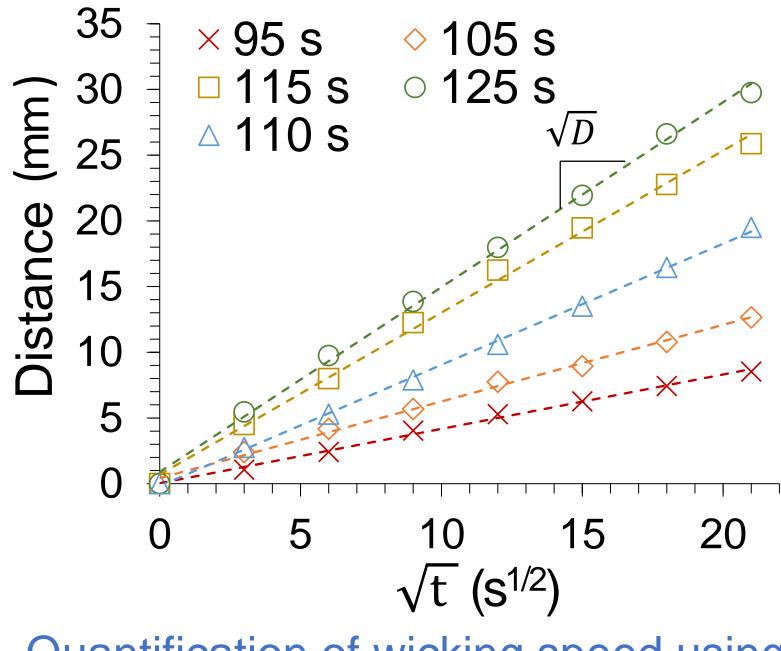
Two-step process to fabricate fully enclosed channels with varying wettability



Fabrication of channels with different wettability in single  $O_2$  plasma step using metal mesh

# **EXPLANATION**





Quantification of wicking speed using Washburn equation